

# Multiple global radiations in tadpole shrimps challenge the concept of ‘living fossils’

Thomas C. Mathers<sup>1</sup>, Robert L. Hammond<sup>2</sup>, Ronald A. Jenner<sup>3</sup>,  
Bernd Hänfling<sup>1</sup> and Africa Gómez<sup>1</sup>

<sup>1</sup> School of Biological, Biomedical and Environmental Sciences, University of Hull, Hull, UK

<sup>2</sup> Department of Biology, University of Leicester, Leicester, UK

<sup>3</sup> Life Sciences Department, The Natural History Museum, London, UK

## ABSTRACT

‘Living fossils’, a phrase first coined by Darwin, are defined as species with limited recent diversification and high morphological stasis over long periods of evolutionary time. Morphological stasis, however, can potentially lead to diversification rates being underestimated. Notostraca, or tadpole shrimps, is an ancient, globally distributed order of branchiopod crustaceans regarded as ‘living fossils’ because their rich fossil record dates back to the early Devonian and their morphology is highly conserved. Recent phylogenetic reconstructions have shown a strong biogeographic signal, suggesting diversification due to continental breakup, and widespread cryptic speciation. However, morphological conservatism makes it difficult to place fossil taxa in a phylogenetic context. Here we reveal for the first time the timing and tempo of tadpole shrimp diversification by inferring a robust multilocus phylogeny of Branchiopoda and applying Bayesian divergence dating techniques using reliable fossil calibrations external to Notostraca. Our results suggest at least two bouts of global radiation in Notostraca, one of them recent, so questioning the validity of the ‘living fossils’ concept in groups where cryptic speciation is widespread.

**Subjects** Evolutionary Studies

**Keywords** Triops, Notostraca, Lepidurus, Biogeography, Diversification, Fossil, Bayesian analysis, ‘Living fossil’, Divergence time

Submitted 20 February 2013

Accepted 14 March 2013

Published 2 April 2013

Corresponding author

Africa Gómez, a.gomez@hull.ac.uk

Academic editor

Keith Crandall

Additional Information and  
Declarations can be found on  
page 8

DOI 10.7717/peerj.62

© Copyright

2013 Mathers et al.

Distributed under

Creative Commons CC-BY 3.0

**OPEN ACCESS**

## INTRODUCTION

There has been much debate about the tempo and mode of the diversification of life (*Eldredge & Gould, 1972; Reznick & Ricklefs, 2009; Rhodes, 1983*). Recently, this debate has been informed by dating using relaxed molecular clocks and diversification analyses; techniques which have revealed disparate patterns of speciation with early bursts (*Burbrink & Pyron, 2010*), recent radiations (*Nagalingum et al., 2011*) and density dependency (*Phillimore & Price, 2008*) being demonstrated. One extreme and often controversial pattern of diversification is found in ‘living fossils’, a concept introduced by Charles Darwin on *the Origin of Species* when dealing with the perplexing nature of the platypus and lungfish, relicts of once diverse groups (*Darwin, 1859*). Since Darwin’s first use, the ‘living fossil’ term has been applied to groups which appear to have diversified little and are morphologically stable over long periods of evolutionary time, with examples including

cycads, tuatara, coelacanths, horseshoe crabs and *Ginkgo biloba*. However, morphological stasis can obscure the patterns of species diversification, and recent time-calibrated phylogenetic analyses of some ‘living fossils’ have indeed revealed that extant species are in fact only recently diverged ([Kano et al., 2012](#); [Nagalingum et al., 2011](#)).

Notostraca, or tadpole shrimps, are an ancient, globally distributed order of branchiopod crustaceans with a rich fossil record dating back to the early Devonian ([Fayers & Trewin, 2002](#)). The order has two extant genera, *Triops* and *Lepidurus*, in the family Triopsidae, with a yet undefined number of species. The nomenclature and systematic position of some ancient extinct Notostraca lineages, is, however, problematic ([Hegna & Dong, 2010](#)). This is partly because tadpole shrimps have maintained an extremely conserved, yet complex, bauplan with extant species indistinguishable from fossils of *Triops* from the Triassic ([Gall & Grauvogel-Stamm, 2005](#); [Gore, 1986](#); [Trusheim, 1938](#)) and of *Lepidurus* in the Jurassic ([Barnard, 1929](#); [Haughton, 1924](#)). This striking morphological conservatism has led them to be referred to as ‘living fossils’ ([Fryer, 1988](#); [King & Hanner, 1998](#); [Mantovani et al., 2008](#)).

Phylogenetic reconstructions of extant Notostraca show a strong biogeographic signal ([Mathers et al., 2013](#); [Vanschoenwinkel et al., 2012](#)). In *Triops*, species complexes are largely restricted to single continents, while *Lepidurus* lineages show high levels of endemism ([Rogers, 2001](#)), patterns that suggest ancient radiation with diversification through continental break-up. However, the extreme morphological conservatism of this order hampers both the taxonomy of extant species and the phylogenetic placement of fossil taxa, with little known about the timing and tempo of notostracan diversification. Genetic analyses have revealed widespread cryptic species ([King & Hanner, 1998](#); [Korn et al., 2010](#); [Korn & Hundsdoerfer, 2006](#); [Macdonald, Sallenave & Cowley, 2011](#); [Vanschoenwinkel et al., 2012](#)), further illustrating the difficulty of inferring past and present diversity. To address this difficulty we infer a robust phylogeny of all known notostracan species from both extant genera and seven branchiopod outgroups. Our analysis uses all available Notostraca sequence data for seven genes, and Bayesian relaxed clock dating techniques, with multiple branchiopod fossil calibrations, to estimate divergence times.

## MATERIALS AND METHODS

### Species delimitation

As Notostraca is known to contain cryptic species complexes (eg. [King & Hanner, 1998](#)), and in order to follow the same criterion for species selection for the multilocus analysis, we delimited species using a generalised mixed Yule coalescent (GMYC) model ([Pons et al., 2006](#)) fitted to an ultrametric phylogeny based on all available cytochrome oxidase I (COI) sequences from GenBank. The 270 sequences were aligned with Muscle ([Edgar, 2004](#)) and phylogeny estimated with BEAST v1.7.4 ([Drummond et al., 2012](#)) under a constant population size coalescent tree model and GTR +  $\Gamma$  substitution model. A strict molecular clock was used with the substitution rate fixed to 1 to provide branch lengths relative to an arbitrary time scale. The MCMC chain was run for 9,000,000 iterations with the first 500,000 iterations removed as burnin. Effective sample sizes (ESS's) of parameters (all greater

than 200) and appropriate burnin were checked using Tracer v1.5 ([Rambaut & Drummond, 2007](#)). From this a maximum clade credibility tree using median heights was made. We then fitted a single threshold GMYC model to the COI tree to delimit species from populations. A total of 34 species of Notostraca were identified in this analysis ([Table S1](#)).

### Multilocus phylogenetic analysis of Branchiopoda

We constructed a multilocus alignment containing representatives of all known species of Notostraca. Single representatives of each phylogenetic species identified by the GMYC analysis were selected for inclusion in our phylogenetic analysis. In addition, four Notostraca lineages (*T. gadensis*, *T. cf. granarius Tunisia*, *L. bilobatus* and *L. cryptus*), which did not have COI data available, but were represented by other genes, were also used in our multilocus phylogenetic analysis. The species status of these lineages has been confirmed in regional studies of cryptic diversity ([King & Hanner, 1998](#); [Korn et al., 2010](#); [Korn & Hundsdoerfer, 2006](#); [Rogers, 2001](#)). We also included seven representatives of the other branchiopod orders so that robust fossil calibrations could be applied for the dating analysis. We included sequences for the genes 12S, 16S, 28S, cytochrome oxidase I (COI), Elongation Factor 1-alpha (EF1), RNA Polymerase II and Glycogen Synthase (see [Table S2](#) for Accession Numbers).

Sequences were aligned using MUSCLE ([Edgar, 2004](#)) with final adjustments by eye. Introns in the nuclear protein coding genes were identified and removed based on alignment with available Notostraca mRNAs. Translation was checked in MEGA 5 ([Tamura et al., 2011](#)). Overall, sequences for 45 taxa (38 notostracan and 7 branchiopod outgroups) were concatenated for analysis with the alignment containing 5793 positions and 52% missing data ([Table S2](#); the alignment file is available in Dryad DOI [10.5061/dryad.77bt2](#)).

Optimum partitioning schemes and substitution models for our phylogenetic analysis and divergence time estimation were identified using PartitionFinder ([Lanfear et al., 2012](#)). PartitionFinder uses a heuristic search algorithm, starting with a fully partitioned analysis (gene and codon position where appropriate), and identifies the best fit partitioning scheme and substitution models based on Bayesian Information Criterion (BIC). Due to the restricted model choice available in RAxML ([Stamatakis, 2006](#)) we conducted separate PartitionFinder analyses for the phylogenetic analysis and divergence time estimation. For the phylogenetic analysis we restricted model choice options to GTR or GTR +  $\Gamma$ , whereas for divergence time estimation we allowed models to be selected from the full suite available in BEAST. We excluded models with proportion of invariant sites (+I) as rate heterogeneity is accounted for by the gamma shape parameter (+ $\Gamma$ ). Optimum partitioning schemes and substitution models for both analyses are given in [Tables S3](#) and [S4](#).

Branchiopod phylogeny was estimated using Bayesian and maximum likelihood (ML) methods with partitions and substitution models set to those identified by PartitionFinder ([Table S3](#)). Bayesian analysis was performed with MrBayes v3.2 ([Ronquist et al., 2012](#)). Model parameters between partitions were unlinked. Two independent MCMC chains

**Table 1** Fossils used to calibrate divergence time analysis in BEAST. Age constraints are treated as hard bounds unless otherwise stated. Node numbers indicate phylogenetic placement of fossil calibrations in Fig. 1.

Node	Fossil taxa	Geological period	Minimum age (Mya)	Maximum age (Mya)	Reference
1	Oldest Bilateria eg. <i>Kimberella</i>	Ediacaran	-	558	<i>Fedonkin, Simonetta &amp; Ivantsov (2007)</i>
1	<i>Rehbachella</i>	Upper Cambrian	-	500 (soft max)	<i>Waloßek (1995)</i>
1	Undescribed anostracan	Base Ordovician	488	-	<i>Harvey, Vélez &amp; Butterfield (2012)</i>
2	<i>Castracollis</i>	Pragian, Early Devonian	410	-	<i>Fayers &amp; Trewin (2002)</i>
3	<i>Ebullitiocaris elatus</i>	Carboniferous	300	-	<i>Womack et al. (2012)</i>
4	<i>Daphnia</i> and <i>Ctenodaphnia</i> sp.	Jurassic/Cretaceous	145	-	<i>Kotov &amp; Taylor (2011)</i>

were run for 10,000,000 iterations each, sampling every 5,000 iterations. The first 25% of each run was discarded as burnin with the remaining samples pooled and used to create a maximum clade credibility tree. Maximum likelihood phylogenetic analysis was performed using RAXMLHPC-PTHREADS v7.0.4 ([Stamatakis, 2006](#)). An initial ML search using GTR +  $\Gamma_4$  was performed onto which 100 rapid bootstraps were drawn.

### Bayesian relaxed clock divergence dating

We estimated Bayesian divergence times with BEAST v1.7.4 ([Drummond et al., 2012](#)) using an uncorrelated lognormal relaxed clock ([Drummond et al., 2006](#)) and a Yule speciation prior. XML files for all BEAST runs were created using BEAUTi v1.7.4 ([Drummond et al., 2012](#)). Topology was constrained to that of the unconstrained RAXML analysis. We used the best fit partitioning scheme identified by PartitionFinder ([Table S4](#)) and estimated substitution model parameters independently for each partition. Initial runs were conducted using substitution models identified by PartitionFinder, however, this resulted in poor mixing of some GTR model parameters for partitions 1, 2 and 5, so subsequent runs were performed using a simpler HKY +  $\Gamma$  model for these partitions.

Five branchiopod fossils representing the oldest known occurrences of their respective crown groups were used to calibrate the molecular clock with minimum age constraints ([Table 1](#)). Lognormal prior distributions were used to specify the level of uncertainty in the placement of these fossil calibrations as they reflect the likely scenario that the true date of divergence of a given node was some time before the earliest known fossil belonging to that clade ([Ho & Phillips, 2009](#)). Lognormal distributions have three parameters – mean, standard deviation and offset. We set the offset to correspond to the minimum age of the node as determined by the fossil record, we then specified mean and standard deviations that resulted in 95% of the distribution falling between the age of the fossil and the age of the next oldest fossil (at a lower taxonomic level) for that group. This gives a prior distribution, which assigns the majority of the probability close to the age of the oldest known fossil, but gives a long tail to account for uncertainty in the proximity of the fossil to the true date of divergence. As Bayesian divergence dating benefits from at

least one maximum age constraint we conservatively constrained the root of the tree to a maximum age of 558 Mya, the age of the oldest bilaterian fossil ([Fedonkin, Simonetta & Ivantsov, 2007](#)).

We ran two independent BEAST MCMC chains for 50,000,000 iterations, sampling every 5000 iterations. 10,000,000 iterations were removed as burnin from each run. Convergence of the two runs and the ESS of parameter estimates (all greater than 250) were assessed using Tracer v1.5 (<http://tree.bio.ed.ac.uk/software/tracer/>). A posterior sample of 8000 trees from one of the runs was used to construct a maximum clade credibility time tree for Notostraca and our selected outgroups. The xml file used to run the BEAST divergence time analysis can be downloaded from Dryad DOI [10.5061/dryad.77bt2](https://doi.org/10.5061/dryad.77bt2).

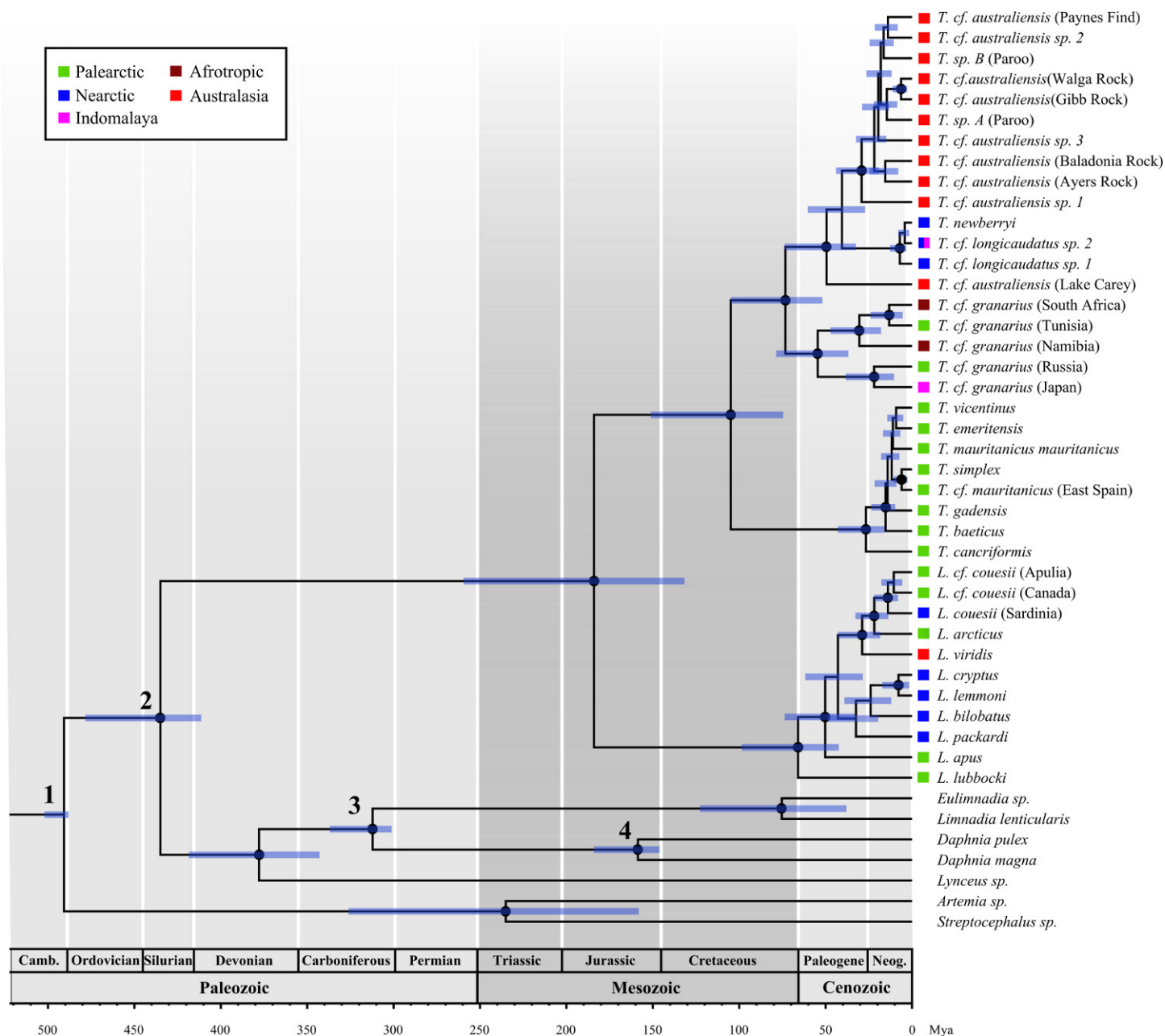
### Diversification analysis

Patterns of diversification through time within Notostraca were investigated using LASER ([Rabosky, 2006](#)) based on the BEAST time tree with outgroups pruned. Using LASER we compared constant rate and variable rate speciation models using likelihood ratio tests ([Table S5](#)).

## RESULTS AND DISCUSSION

The most complete taxon sampling to date coupled with the inclusion of multiple nuclear and mitochondrial markers allowed us to generate a robust phylogeny of 38 extant Notostraca species. ML and Bayesian inference gave highly congruent phylogenetic trees with most branches highly supported ([Figs. S1 and S2](#)). The recovered relationships between branchiopod orders are in agreement with recently published arthropod phylogenies ([Regier et al., 2010](#); [von Reumont et al., 2012](#)), providing a solid platform for divergence dating analysis.

Our robust time-calibrated phylogeny of Branchiopoda ([Fig. 1](#)) clearly shows that Notostraca has a pattern of diversity incompatible with Darwin's original usage of the term 'living fossil' as relics of once diverse groups, and importantly reveals cryptic patterns of diversification. Our analysis – using outgroup fossil calibrations – estimates an ancient divergence of *Triops* and *Lepidurus* during the Jurassic, 184 Mya (95% confidence interval 132–259 Mya), which agrees with the earliest fossils assigned to *Lepidurus* ([Barnard, 1929](#); [Gand, Garric & Lapeyrie, 1997](#)), and with a sister relationship of Notostraca to the extinct order Kazacharthra of the Late Triassic/Early Jurassic ([Olesen, 2009](#)). This initial radiation of extant Notostraca was not, however, due to continental break up as the timing and pattern of diversification within the genera substantially postdates the break-up of Pangaea 160–138 Mya ([Scotese, 2001](#)). Furthermore, the current species distributions of *Triops* and *Lepidurus* are likely to have resulted from a second global radiation of the order, possibly following considerable levels of extinction. We conclude this because fossil Notostraca, attributed to *Triops* and *Lepidurus*, have been found in modern day North and South America, Europe, Africa and Antarctica, implying a global distribution by the early Jurassic ([Gall & Grauvogel-Stamm, 2005](#); [Gand, Garric & Lapeyrie, 1997](#); [Garrouste, Nel & Gand, 2009](#); [Gore, 1986](#); [Haughton, 1924](#); [Trusheim, 1938](#)). Yet, our LASER analysis shows a

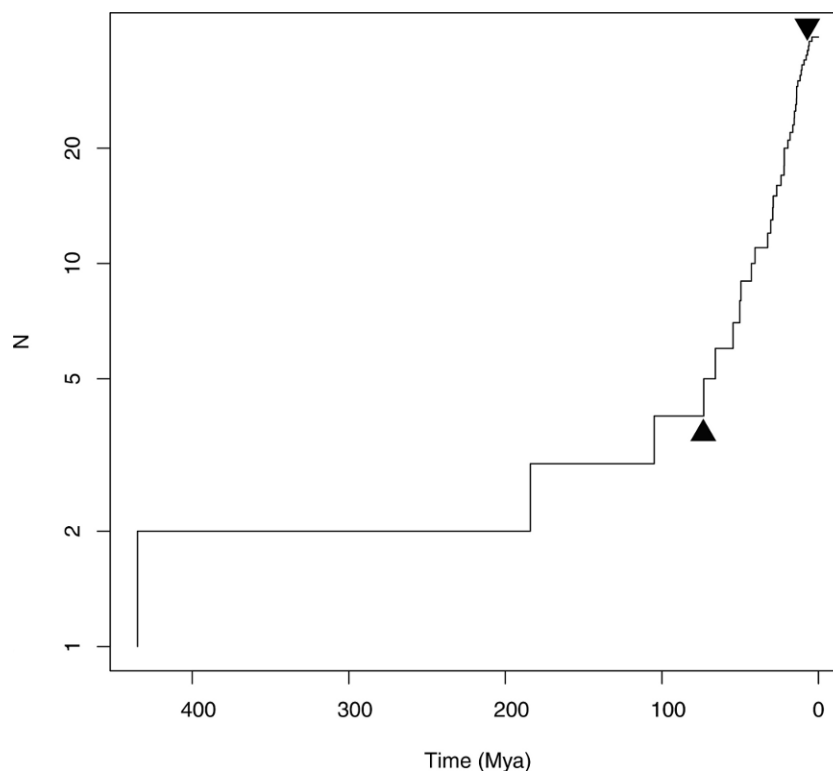


**Figure 1** Time calibrated phylogeny of 38 notostracan species and seven branchiopod outgroups. Numbers at nodes correspond to the fossil calibrations given in Table 1. Nodes with black circles have ML Bootstrap support values greater than 70 and posterior probabilities greater than 95 from the RAXML and MrBayes analyses respectively. Error bars show the 95% confidence intervals of divergence times. Colour coded squares show the known geographic distribution of each species.

significant increase in the rate of diversification of Notostraca about 73 Mya (Fig. 2), close to the time of the K-Pg mass extinction event. It is this second radiation that resulted in the current global distribution of extant *Triops* and *Lepidurus*.

The almost synchronous radiation of *Triops* and *Lepidurus* (Fig. 1) suggests that a common factor may have triggered diversification of the two genera. The diversification





**Figure 2** Diversification of Notostraca through time. Arrows indicate the timing and direction of shifts in rate of diversification inferred by LASER.  $N$  is the number of species. The best fit ML model of diversification identified three distinct rates of diversification during the evolutionary history of Notostraca with an increase in speciation rate 73 Mya followed by a decrease 6 Mya.

of modern birds – widely involved in dispersal in aquatic invertebrates (Green *et al.*, 2005; van Leeuwen *et al.*, 2012) – coincided with the initiation of the notostracan radiation (Pacheco *et al.*, 2011), and may have facilitated the long distance dispersal and subsequent diversification of Notostraca. Indeed, the geographical distribution of extant taxa (Fig. 1) suggests several instances of intercontinental dispersal. For example, the colonisation of North America from Australia could have resulted from dispersal events during bird migration. Such long distance dispersal and colonisation events might also have been facilitated by the flexible nature of sexual systems found within Notostraca (Mathers *et al.*, 2013). Indeed, the evolution of androdioecy – a sexual system where males and hermaphrodites coexist (Weeks, 2012; Zierold, Hanfling & Gómez, 2007; Zierold *et al.*, 2009) – from gonochorism appears to have favoured postglacial recolonisation in the species *Triops cancriformis* (Zierold, Hanfling & Gómez, 2007).

The concept of ‘living fossils’ has been a controversial one as it has often been interpreted to imply a lack of evolutionary change, even against evidence of molecular evolutionary change (Avisé, Nelson & Sugita, 1994; Casane & Laurenti, 2013). Our divergence dating analysis has shown that tadpole shrimps can be regarded as ‘living fossils’ only on the grounds of morphological conservatism, not on the basis of limited diversification or relict status. Instead, throughout their long evolutionary history,

notostracans have undergone multiple global radiations and high species turnover. Recent, time calibrated, phylogenetic analysis of other traditional ‘living fossils’ such as cycads (*Nagalingum et al., 2011*), nautiloids (*Wray et al., 1995*) horseshoe crabs (*Obst et al., 2012*) and monoplacophorans (*Kano et al., 2012*), have also revealed that extant species are more recently diverged than suggested by fossil data alone. We therefore caution against drawing conclusions about patterns of diversification based on fossil data alone in groups where widespread morphological conservatism may obscure rampant cryptic speciation. Furthermore, our results help clarify the term ‘living fossils’, putting important questions into focus. Namely, is such morphological conservatism, in the face of evolutionarily recent diversification and radiation, best accounted for by unchanging selection or by developmental genetic constraints?

## ACKNOWLEDGEMENTS

This work was part of TCM’s Ph.D. We thank Chris Venditti for his help with the analyses, and Dave Lunt and Steve Moss for bioinformatics support. Domino Joyce read and provided constructive comments in a previous version of the manuscript.

## ADDITIONAL INFORMATION AND DECLARATIONS

### Funding

This work was part of TCM’s Ph.D., funded by a NERC CASE Studentship NE/G012318/1. AG was supported by a NERC Advanced fellowship NE/B501298/1. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

### Grant Disclosures

The following grant information was disclosed by the authors:

NERC CASE Studentship: NE/G012318/1.

NERC Advanced fellowship: NE/B501298/1.

### Competing Interests

There are no Competing Interests to declare.

### Author Contributions

- Thomas C. Mathers conceived and designed the experiments, performed the experiments, analyzed the data, wrote the paper.
- Robert L. Hammond, Ronald A. Jenner, Bernd Hänfling and Africa Gómez conceived and designed the experiments, wrote the paper.

### Data Deposition

The following information was supplied regarding the deposition of related data:

We will deposit alignments and files used to produce the results in Dryad.



## Supplemental Information

Supplemental information for this article can be found online at <http://dx.doi.org/10.7717/peerj.62>.

## REFERENCES

- Avisé JC, Nelson WS, Sugita H. 1994.** A speciation history of “living fossils”: molecular evolutionary patterns in horseshoe crabs. *Evolution* **48**:1986–2001 DOI [10.2307/2410522](https://doi.org/10.2307/2410522).
- Barnard KH. 1929.** A revision of the South African branchiopoda phyllopoda. *Annals of the South African Museum* **29**:181–272.
- Burbrink FT, Pyron RA. 2010.** How does ecological opportunity influence rates of speciation, extinction, and morphological diversification in New World ratsnakes (tribe Lampropeltini)? *Evolution* **64**:934–943 DOI [10.1111/j.1558-5646.2009.00888.x](https://doi.org/10.1111/j.1558-5646.2009.00888.x).
- Casane D, Laurenti P. 2013.** Why coelacanths are not ‘living fossils’. *BioEssays* **35**(4):332–338 DOI [10.1002/bies.201200145](https://doi.org/10.1002/bies.201200145).
- Darwin C. 1859.** *On the origin of species by means of natural selection*. London: Murray.
- Drummond AJ, Ho SYW, Phillips MJ, Rambaut A. 2006.** Relaxed phylogenetics and dating with confidence. *PLoS Biology* **4**:e88 DOI [10.1371/journal.pbio.0040088](https://doi.org/10.1371/journal.pbio.0040088).
- Drummond AJ, Suchard MA, Xie D, Rambaut A. 2012.** Bayesian phylogenetics with BEAUti and the BEAST 1.7. *Molecular Biology and Evolution* **29**:1969–1973 DOI [10.1093/molbev/mss075](https://doi.org/10.1093/molbev/mss075).
- Edgar RC. 2004.** MUSCLE: multiple sequence alignment with high accuracy and high throughput. *Nucleic Acids Research* **32**:1792–1797 DOI [10.1093/nar/gkh340](https://doi.org/10.1093/nar/gkh340).
- Eldredge N, Gould SJ. 1972.** Punctuated equilibria: an alternative to phyletic gradualism. *Models in Paleobiology* **82**:115.
- Fayers SR, Trewin NH. 2002.** A new crustacean from the Early Devonian Rhynie chert, Aberdeenshire, Scotland. *Earth and Environmental Science Transactions of the Royal Society of Edinburgh* **93**:355–382 DOI [10.1017/S026359330000047X](https://doi.org/10.1017/S026359330000047X).
- Fedonkin MA, Simonetta A, Ivantsov AY. 2007.** New data on *Kimberella*, the Vendian mollusc-like organism (White Sea region, Russia): palaeoecological and evolutionary implications. London: Geological Society, **286**:157–179. Special Publications.
- Fryer G. 1988.** Studies on the functional-morphology and biology of the Notostraca (Crustacea, Branchiopoda). *Philosophical Transactions of the Royal Society of London Series B-Biological Sciences* **321**:27–142 DOI [10.1098/rstb.1988.0091](https://doi.org/10.1098/rstb.1988.0091).
- Gall J-C, Grauvogel-Stamm L. 2005.** The early middle Triassic ‘Grès à Voltzia’ formation of eastern France: a model of environmental refugium. *Paléontologie générale* **4**:569–581.
- Gand G, Garric J, Lapeyrie J. 1997.** Biocénoses à triopsidés (Crustacea, Branchiopoda) du Permien du bassin de Lodève (France). *Geobios* **30**:673–700 DOI [10.1016/S0016-6995\(97\)80157-X](https://doi.org/10.1016/S0016-6995(97)80157-X).
- Garrouste R, Nel A, Gand G. 2009.** New fossil arthropods (Notostraca and Insecta: Syntonopterida) in the continental Middle Permian of Provence (Bas-Argens Basin, France). *Comptes Rendus Palevol* **8**:49–57 DOI [10.1016/j.crpv.2008.11.008](https://doi.org/10.1016/j.crpv.2008.11.008).
- Gore PJW. 1986.** Triassic notostracans in the newark basin, New Jersey and Northern Virginia. *Journal of Paleontology* **60**:1086–1096.
- Green AJ, Sánchez MI, Amat F, Figuerola J, Hontela A. 2005.** Dispersal of invasive and native brine shrimps *Artemia* (Anostraca) via waterbirds. *Limnology and Oceanography* **50**:737–742 DOI [10.4319/lo.2005.50.2.0737](https://doi.org/10.4319/lo.2005.50.2.0737).

- Harvey THP, Vélez MI, Butterfield NJ. 2012. Exceptionally preserved crustaceans from western Canada reveal a cryptic Cambrian radiation. *Proceedings of the National Academy of Sciences* **109**:1589–1594 DOI [10.1073/pnas.1115244109](https://doi.org/10.1073/pnas.1115244109).
- Haughton S. 1924. The fauna and stratigraphy of the Stormberg *Museum* **12**:323–497.
- Hegna TA, Dong REN. 2010. Two new “Notostracans”, *Chenops* gen. nov. and *Jenolops* gen. nov. (Crustacea: Branchiopoda: Notostraca) from the Yixian Formation, Northeastern China. *Acta Geologica Sinica* **84**:886–894 (English Edition) DOI [10.1111/j.1755-6724.2010.00232.x](https://doi.org/10.1111/j.1755-6724.2010.00232.x).
- Ho SYW, Phillips MJ. 2009. Accounting for calibration uncertainty in phylogenetic estimation of evolutionary divergence times. *Systematic Biology* **58**:267–280 DOI [10.1093/sysbio/syp035](https://doi.org/10.1093/sysbio/syp035).
- Kano Y, Kimura S, Kimura T, Warén A. 2012. Morphological conservatism or recent diversification? DOI [10.1111/j.1463-6409.2012.00550.x](https://doi.org/10.1111/j.1463-6409.2012.00550.x).
- King JL, Hanner R. 1998. Cryptic species in taxonomic and phylogenetic relationships within the genus *Lepidurus* (Crustacea: Notostraca) in North America. *Molecular Phylogenetics and Evolution* **10**:23–36 DOI [10.1006/mpev.1997.0470](https://doi.org/10.1006/mpev.1997.0470).
- Korn M, Green AJ, Machado M, Garcia-de-Lomas J, Cristo M, da Fonseca LC, Frisch D, Perez-Bote JL, Hundsdoerfer AK. 2010. Phylogeny, molecular ecology and taxonomy of southern Iberian lineages of *Triops mauritanicus* (Crustacea: Notostraca). *Organisms Diversity and Evolution* **10**:409–440 DOI [10.1007/s13127-010-0026-y](https://doi.org/10.1007/s13127-010-0026-y).
- Korn M, Hundsdoerfer AK. 2006. Evidence for cryptic species in the tadpole shrimp *Triops granarius* (Lucas, 1864) (Crustacea: Notostraca). *Zootaxa* **1257**:57–68.
- Kotov AA, Taylor DJ. 2011. Mesozoic fossils (>145 Mya) suggest the antiquity of the subgenera of *Daphnia* and their coevolution with chaoborid predators. *BMC Evolutionary Biology* **11**:129 DOI [10.1186/1471-2148-11-129](https://doi.org/10.1186/1471-2148-11-129).
- Lanfear R, Calcott B, Ho SYW, Guindon S. 2012. Partitionfinder: combined selection of partitioning schemes and substitution models for phylogenetic analyses. *Molecular Biology and Evolution* **29**:1695–1701 DOI [10.1093/molbev/mss020](https://doi.org/10.1093/molbev/mss020).
- Macdonald KS, Sallenave R, Cowley DE. 2011. Morphological and genetic variation in *Triops* (Branchiopoda: Notostraca) from ephemeral waters of the Northern Chihuahuan desert of North America. *Journal of Crustacean Biology* **31**:468–484 DOI [10.1651/10-3411.1](https://doi.org/10.1651/10-3411.1).
- Mantovani B, Cesari M, Luchetti A, Scanabissi F. 2008. Mitochondrial and nuclear DNA variability in the living fossil *Triops cancriformis* (Bosc, 1801) (Crustacea, Branchiopoda, Notostraca). *Heredity* **100**:496–505 DOI [10.1038/hdy.2008.3](https://doi.org/10.1038/hdy.2008.3).
- Mathers TC, Hammond RL, Jenner RA, Zierold T, Hänfling B, Gómez A. 2013. High lability of sexual system over 250 million years of evolution in morphologically conservative tadpole shrimps. *BMC Evolutionary Biology* **13**:30 DOI [10.1186/1471-2148-13-30](https://doi.org/10.1186/1471-2148-13-30).
- Nagalingum NS, Marshall CR, Quental TB, Rai HS, Little DP, Mathews S. 2011. Recent synchronous radiation of a living fossil. *Science* **334**:796–799 DOI [10.1126/science.1209926](https://doi.org/10.1126/science.1209926).
- Obst M, Faurby S, Bussarawit S, Funch P. 2012. Molecular phylogeny of extant horseshoe crabs (Xiphosura, Limulidae) indicates Paleogene diversification of Asian species. *Molecular Phylogenetics and Evolution* **62**:21–26 DOI [10.1016/j.ympev.2011.08.025](https://doi.org/10.1016/j.ympev.2011.08.025).
- Olesen J. 2009. Phylogeny of Branchiopoda (Crustacea)—character evolution and contribution of uniquely preserved fossils. *Arthropod Systematics and Phylogeny* **67**:3–39.

- Pacheco MA, Battistuzzi FU, Lentino M, Aguilar R, Kumar S, Escalante AA. 2011. Evolution of modern birds revealed by Mitogenomics: timing the radiation and origin of major orders. *Molecular Biology and Evolution* 28:1927–1942 DOI 10.1093/molbev/msr014.
- Phillimore AB, Price TD. 2008. Density-dependent cladogenesis in birds. *PLoS Biology* 6:e71 DOI 10.1371/journal.pbio.0060071.
- Pons J, Barraclough TG, Gomez-Zurita J, Cardoso A, Duran DP, Hazell S, Kamoun S, Sumlin WD, Vogler AP. 2006. Sequence-based species delimitation for the DNA taxonomy of undescribed insects. *Systematic Biology* 55:595–609 DOI 10.1080/10635150600852011.
- Rabosky DL. 2006. LASER: a maximum likelihood toolkit for detecting temporal shifts in diversification rates from molecular phylogenies. *Evol Bioinformatics Online* 2:273–276.
- Rambaut A, Drummond A. 2007. Tracer v1. 4 [upated to v1. 5].
- Regier JC, Shultz JW, Zwick A, Hussey A, Ball B, Wetzer R, Martin JW, Cunningham CW. 2010. Arthropod relationships revealed by phylogenomic analysis of nuclear protein-coding sequences. *Nature* 463:1079–1083 DOI 10.1038/nature08742.
- Reznick DN, Ricklefs RE. 2009. Darwin’s bridge between microevolution and macroevolution. *Nature* 457:837–842 DOI 10.1038/nature07894.
- Rhodes FHT. 1983. Gradualism, punctuated equilibrium and the origin of species. *Nature* 305:269–272 DOI 10.1038/305269a0.
- Rogers DC. 2001. Revision of the nearctic *Lepidurus* (Notostraca). *Journal of Crustacean Biology* 21:991–1006 DOI 10.1651/0278-0372(2001)021[0991:ROTNLN]2.0.CO;2.
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP. 2012. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology* 61:539–542 DOI 10.1093/sysbio/sys029.
- Scotese CR. 2001. Atlas of earth history: University of Texas at Arlington. Department of Geology. PALEOMAP Project.
- Stamatakis A. 2006. RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* 22:2688–2690 DOI 10.1093/bioinformatics/btl446.
- Tamura K, Peterson D, Peterson N, Stecher G, Nei M, Kumar S. 2011. MEGA5: molecular evolutionary genetics analysis using maximum likelihood, evolutionary distance, and maximum parsimony methods. *Molecular Biology and Evolution* 28:2731–2739 DOI 10.1093/molbev/msr121.
- Trusheim F. 1938. Triopsiden (Crust. Phyll.) aus dem Keuper Frankens. *Paläontologische Zeitschrift* 19:198–216.
- van Leeuwen CHA, van der Velde G, van Groenendaal JM, Klaassen M. 2012. Gut travellers: internal dispersal of aquatic organisms by waterfowl. *Journal of Biogeography* 39:2031–2040 DOI 10.1111/jbi.12004.
- Vanschoenwinkel B, Pinceel T, Vanhove MPM, Denis C, Jocque M, Timms BV, Brendonck L. 2012. Toward a global phylogeny of the “living fossil” crustacean order of the Notostraca. *PLoS ONE* 7:e34998 DOI 10.1371/journal.pone.0034998.
- von Reumont BM, Jenner RA, Wills MA, Dell’Ampio E, Pass G, Ebersberger I, Meyer B, Koenemann S, Iliffe TM, Stamatakis A, Niehuis O, Meusemann K, Misof B. 2012. Pancrustacean phylogeny in the light of new phylogenomic data: support for Remipedia

- as the possible sister group of hexapoda. *Molecular Biology and Evolution* **29**:1031–1045 DOI [10.1093/molbev/msr270](https://doi.org/10.1093/molbev/msr270).
- Waloßek D. 1995.** The upper cambrian *Rehbachella*, its larval development, morphology and significance for the phylogeny of branchiopoda and crustacea. *Hydrobiologia* **298**:1–13 DOI [10.1007/BF00033797](https://doi.org/10.1007/BF00033797).
- Weeks SC. 2012.** The role of androdioecy and gynodioecy in mediating evolutionary transitions between dioecy and hermaphroditism in the Animalia. *Evolution* **66**:3670–3686 DOI [10.1111/j.1558-5646.2012.01714.x](https://doi.org/10.1111/j.1558-5646.2012.01714.x).
- Womack T, Slater BJ, Stevens LG, Anderson LI, Hilton J. 2012.** First cladoceran fossils from the carboniferous: palaeoenvironmental and evolutionary implications. *Palaeogeography, Palaeoclimatology, Palaeoecology* **344–345**:39–48 DOI [10.1016/j.palaeo.2012.05.012](https://doi.org/10.1016/j.palaeo.2012.05.012).
- Wray CG, Landman NH, Saunders WB, Bonacum J. 1995.** Genetic divergence and geographic diversification in *Nautilus*. *Paleobiology* **21**:220–228.
- Zierold T, Hänfling B, Gómez A. 2007.** Recent evolution of alternative reproductive modes in the ‘living fossil’ *Triops cancriformis*. *BMC Evolutionary Biology* **7**:161 DOI [10.1186/1471-2148-7-161](https://doi.org/10.1186/1471-2148-7-161).
- Zierold T, Montero-Pau J, Hänfling B, Gómez A. 2009.** Sex ratio, reproductive mode and genetic diversity in *Triops cancriformis*. *Freshwater Biology* **54**:1392–1405 DOI [10.1111/j.1365-2427.2009.02191.x](https://doi.org/10.1111/j.1365-2427.2009.02191.x).

---

## Author Queries

*Journal:* PEERJ

*Article id:* 62

*Author:* Mathers et al.

*Title:* Multiple global radiations in tadpole shrimps challenge the concept of 'living fossils'

---

### Q1 (Page 9)

Complete reference informations are updated for the reference(s): Casane and Laurenti, 2013; von Reumont et al., 2012. Please check, and correct if necessary.

### Q2 (Page 9)

Please provide page extents for the reference(s): Eldredge and Gould, 1972; Kotov and Taylor, 2011; Mathers et al. 2013; Zierold et al. 2007.

### Q3 (Page 9)

Please check page number given in references Gall and Grauvogel-Stamm, 2005 and Regier, 2010.

### Q4 (Page 11)

Please check the journal title given in Rabosky, 20



### Q5 (Page 11)

Please provide complete reference information for the reference(s): Rambaut and Drummond, 2007.

---